PUBLIC HEALTH GOALS FOR CHEMICALS IN DRINKING WATER

1,1,2-TRICHLOROETHANE

March 2006

Governor of the State of California Arnold Schwarzenegger

Secretary for Environmental Protection California Environmental Protection Agency Alan C. Lloyd, Ph.D.

Director Office of Environmental Health Hazard Assessment Joan E. Denton, Ph.D.



Public Health Goal for 1,1,2-TRICHLOROETHANE in Drinking Water

Prepared by

Office of Environmental Health Hazard Assessment California Environmental Protection Agency

Pesticide and Environmental Toxicology Branch Anna M. Fan, Ph.D., Chief

> Deputy Director for Scientific Affairs George V. Alexeeff, Ph.D.

> > **March 2006**

LIST OF CONTRIBUTORS

PHG PROJECT MANAGEMENT

REPORT PREPARATION

SUPPORT

Project Director Anna Fan, Ph.D.

Author Thomas Parker, M.S.

Primary Reviewer John Faust, Ph.D.

Final Reviewers Anna Fan, Ph.D. George Alexeeff, Ph.D. Robert Howd, Ph.D. Administrative Support Genevieve Vivar Sharon Davis Hermelinda Jimenez

Library Support Charleen Kubota, M.L.S.

> *Web site Posting* Laurie Monserrat

We thank the U.S. Environmental Protection Agency (Office of Water; National Center for Environmental Assessment) and the faculty members of the University of California with whom the Office of Environmental Health Hazard Assessment contracted through the University of California Office of the President for their peer reviews of the public health goal documents, and gratefully acknowledge the comments received from all interested parties.

1,1,2-Trichloroethane in Drinking Water California Public Health Goal ii

rector Author

PHG Program Leader Robert Howd, Ph.D.

Comment Coordinator Catherine Caraway, B.S.

Revisions/Responses Robert Howd, Ph.D.

PREFACE

Drinking Water Public Health Goal Pesticide and Environmental Toxicology Branch Office of Environmental Health Hazard Assessment California Environmental Protection Agency

This Public Health Goal (PHG) technical support document provides information on health effects from contaminants in drinking water. PHGs are developed for chemical contaminants based on the best available toxicological data in the scientific literature. These documents and the analyses contained in them provide estimates of the levels of contaminants in drinking water that would pose no significant health risk to individuals consuming the water on a daily basis over a lifetime.

The California Safe Drinking Water Act of 1996 (Health and Safety Code, Section 116365) requires the Office of Environmental Health Hazard Assessment (OEHHA) to perform risk assessments and adopt PHGs for contaminants in drinking water based exclusively on public health considerations. The Act requires that PHGs be set in accordance with the following criteria:

- 1. PHGs for acutely toxic substances shall be set at levels at which no known or anticipated adverse effects on health will occur, with an adequate margin of safety.
- 2. PHGs for carcinogens or other substances that may cause chronic disease shall be based solely on health effects and shall be set at levels that OEHHA has determined do not pose any significant risk to health.
- 3. To the extent the information is available, OEHHA shall consider possible synergistic effects resulting from exposure to two or more contaminants.
- 4. OEHHA shall consider potential adverse effects on members of subgroups that comprise a meaningful proportion of the population, including but not limited to infants, children, pregnant women, the elderly, and individuals with a history of serious illness.
- 5. OEHHA shall consider the contaminant exposure and body burden levels that alter physiological function or structure in a manner that may significantly increase the risk of illness.
- 6. OEHHA shall consider additive effects of exposure to contaminants in media other than drinking water, including food and air, and the resulting body burden.
- 7. In risk assessments that involve infants and children, OEHHA shall specifically assess exposure patterns, special susceptibility, multiple contaminants with toxic mechanisms in common, and the interactions of such contaminants.

- 8. In cases of insufficient data for OEHHA to determine a level that creates no significant risk, OEHHA shall set the PHG at a level that is protective of public health with an adequate margin of safety.
- 9. In cases where scientific evidence demonstrates that a safe dose response threshold for a contaminant exists, then the PHG should be set at that threshold.
- 10. The PHG may be set at zero if necessary to satisfy the requirements listed above in items seven and eight.
- 11. PHGs adopted by OEHHA shall be reviewed at least once every five years and revised as necessary based on the availability of new scientific data.

PHGs adopted by OEHHA are for use by the California Department of Health Services (DHS) in establishing primary drinking water standards (State Maximum Contaminant Levels, or MCLs). Whereas PHGs are to be based solely on scientific and public health considerations without regard to economic cost considerations or technical feasibility, drinking water standards adopted by DHS are to consider economic factors and technical feasibility. Each primary drinking water standard adopted by DHS shall be set at a level that is as close as feasible to the corresponding PHG, placing emphasis on the protection of public health. PHGs established by OEHHA are not regulatory in nature and represent only non-mandatory goals. By state and federal law, MCLs established by DHS must be at least as stringent as the federal MCL, if one exists.

PHG documents are used to provide technical assistance to DHS, and they are also informative reference materials for federal, state and local public health officials and the public. While the PHGs are calculated for single chemicals only, they may, if the information is available, address hazards associated with the interactions of contaminants in mixtures. Further, PHGs are derived for drinking water only and are not intended to be utilized as target levels for the contamination of other environmental media.

Additional information on PHGs can be obtained at the OEHHA Web site at www.oehha.ca.gov.

TABLE OF CONTENTS

LIST OF CONTRIBUTORS	II
PREFACE	111
TABLE OF CONTENTS	V
PUBLIC HEALTH GOAL FOR 1,1,2-TRICHLOROETHANE IN DI WATER	
SUMMARY	1
INTRODUCTION	2
CHEMICAL PROFILE	2
Chemical Identity	2
Physical and Chemical Properties	
Production and Uses	4
ENVIRONMENTAL OCCURRENCE AND HUMAN EXPOSURE	4
Air	4
Soil	4
Water	4
METABOLISM AND PHARMACOKINETICS	5
Absorption	5
Distribution	5
Metabolism	5
Excretion	6
TOXICOLOGY	6
Toxicological Effects in Animals	6
Acute Toxicity	6
Subchronic Toxicity	7
Genetic Toxicity	8
Developmental and Reproductive Toxicity	10
Immunotoxicity	10

Neurotoxicity	10
Chronic Toxicity	10
Carcinogenicity	11
Oral Exposure	11
Subcutaneous Exposure	
Transgenic Mouse Studies	
Toxicological Effects in Humans	
Acute Toxicity	
Subchronic Toxicity	
Genetic Toxicity	13
Developmental and Reproductive Toxicity	13
Immunotoxicity	13
Neurotoxicity	13
Chronic Toxicity	
Carcinogenicity	14
DOSE-RESPONSE ASSESSMENT	14
Noncarcinogenic Effects	14
Carcinogenic Effects	14
CALCULATION OF PHG	15
Exposure Considerations	15
Noncarcinogenic Effects	15
Carcinogenic Effects	16
RISK CHARACTERIZATION	17
OTHER REGULATORY STANDARDS	
REFERENCES	

PUBLIC HEALTH GOAL FOR 1,1,2-TRICHLOROETHANE IN DRINKING WATER

SUMMARY

1,1,2-Trichloroethane (1,1,2-TCA) is a halogenated aliphatic hydrocarbon. It is a nonflammable liquid with a pleasant odor, insoluble in water, and miscible with alcohol, ether, and many other organic liquids. 1,1,2-Trichloroethane is used in manufacturing 1,1-dichloroethene; as a solvent for fats, waxes, natural resins, and alkaloids; and in other organic syntheses. Small amounts of 1,1,2-TCA are formed during chlorination of drinking water.

Limited human data are available on the toxicity of 1,1,2-TCA. It is rapidly absorbed by inhalation, and can exert a narcotic effect. 1,1,2-Trichloroethane is irritating to the eyes, nose, and respiratory tract. When the liquid solvent comes in contact with skin, it may cause cracking and erythema. Animal studies showed nervous system effects such as excitation and sleepiness, and at high levels, toxicity to the liver and kidneys.

No epidemiologic studies are available to address adequately the potential for carcinogenicity of 1,1,2-TCA in humans. In a rodent bioassay of 1,1,2-TCA by gavage, no effects on tumor development were noted in rats, while treated mice had significantly increased incidences of hepatocellular carcinomas and adrenal pheochromocytomas. 1,1,2-Trichloroethane is negative in genotoxicity studies. The carcinogenic potential of 1,1,2-TCA is a concern, and the data are adequate to use for estimating a human health-protective level of 1,1,2-TCA in drinking water. 1,1,2-TCA is listed as "known to the state to cause cancer" by its synonym, vinyl trichloride, under California's Safe Drinking Water and Toxic Enforcement Act of 1986 (Proposition 65). The U.S. EPA classifies 1,1,2-TCA as a possible human carcinogen (group C), and provides a cancer potency factor based upon the mouse tumor data (U.S. EPA, 2004). The International Agency for Research on Cancer (IARC) has classified 1,1,2-TCA as "not classifiable as to its carcinogenicity to humans" (Group 3) based upon limited evidence in experimental animals and the absence of epidemiological evidence (IARC, 1999).

The hereby establish a Public Health Goal (PHG) for 1,1,2-TCA of 0.0003 mg/L, based on the 95 percent upper confidence bound of the slope of the fit by the linearized multistage model to the carcinogenicity dose-response data, calculated for a one in one million cancer risk with a lifetime of exposure. This value is considerably lower than the current California and federal Maximum Contaminant Level (MCL) of 0.005 mg/L, based on non-cancer toxic effects and considerations of analytical feasibility. The basis of the PHG is positive evidence of carcinogenicity in oral cancer bioassays in mice performed by the National Cancer Institute (NCI, 1978). NCI examined the effects of technical grade 1,1,2-TCA administered orally in corn oil to mice and rats. Groups of 50 male and female B6C3F₁ mice were treated via gavage five days/week for 78 weeks, with time-weighted average doses of 195 and 390 mg/kg-d, respectively, for the low and high dose groups for both sexes over the course of the dosing. In the high dose males, 37/49 (76 percent), and in the high dose females, 40/45 (89 percent), developed hepatocellular

carcinoma. The results in rats were negative. Both OEHHA and U.S. EPA have judged these data in mice adequate for estimating a cancer potency value (OEHHA, 1999; U.S. EPA, 2004). A positive association between administration of 1,1,2-TCA and the incidence of pheochromocytoma of the adrenal gland was also observed in the high dose female mice (NCI, 1978).

INTRODUCTION

The purpose of this document is to develop a PHG for the chlorinated hydrocarbon solvent 1,1,2-trichloroethane (1,1,2-TCA) in drinking water. This task involves performing an updated literature review and a detailed risk assessment. A primary objective was to determine if there is a more appropriate toxicological study or a better method of calculating a public-health protective level for 1,1,2-TCA in drinking water than was used in the prior risk assessment used to develop the existing California MCL. This document is not intended to provide an exhaustive review of all aspects of the use and toxicology of 1,1,2-TCA, but to concentrate on the lowest-dose for the critical toxic effects that may contribute to public health concern from exposure to the solvent in drinking water.

U.S. EPA proposed a federal Maximum Contaminant Level Goal (MCLG) of 0.003 mg/L for 1,1,2-TCA on the basis of liver, kidney or immune system toxicity in 1990 (U.S. EPA, 1990, 2001a). An MCL of 0.005 mg/L was set based on analytical feasibility at that time, and the two criteria were finalized in 1992 (U.S. EPA, 1992, 2002). The California MCL was subsequently revised to match the federal MCL (see DHS, 2002).

CHEMICAL PROFILE

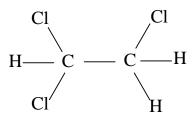
Chemical Identity

The structure, CAS registry number, and chemical formula are given below, as well as various names used to identify this solvent.

Parameter	Property or Value
Chemical name	1,1,2-trichloroethane
Synonyms	Ethane trichloride, vinyl trichloride, β- trichloroethane, 1,2,2-trichloroethane
Registered trade name	β-T, Cement T-339
Chemical formula	C ₂ H ₃ Cl ₃
CAS Registry Number	79-00-5

Table 1. Chemical Identity of 1,1,2-TCA

Figure 1. Chemical structure of 1,1,2-TCA



Physical and Chemical Properties

1,1,2-Trichloroethane is a volatile, lipophilic chlorinated solvent. 1,1,2-Trichloroethane can be sensitively detected by a variety of instruments including mass spectrometry and an electron capture detector. No matter which detection instrument is used, the first step of most analyses is to separate 1,1,2-TCA from the water and non-volatile chemicals in the water by purging the volatile chemicals from the water with an inert gas and trapping them on a solid absorbent (Cleseri *et al.*, 1989). Physical and chemical properties relevant to assessing human exposure to 1,1,2-TCA are provided in Table 2.

Property	Value or Information	References
Molecular weight	133.4	NIOSH, 1994
Color	Colorless	NIOSH, 1994
Physical state	Liquid	NIOSH, 1994
Odor	Chloroform-like, sweet	NIOSH, 1994
Melting point	-36 °C	U.S. EPA, 2001b
Boiling point	113.8 °C	U.S. EPA, 2001b
Flash point	N/A	NIOSH, 1994
Flammability limits	UEL 15.5%, LEL 6%	NIOSH, 1994
Solubility, Water	4.4 g/L at 20 °C	U.S. EPA, 2001b
Specific gravity, density	1.4 at 20 °C	U.S. EPA, 2001b
Octanol/water log Kow	2.17	U.S. EPA, 2001b
Vapor pressure	23 mm Hg at 25 °C	U.S. EPA, 2001b
Conversion factor	$1 \text{ ppm} = 5.5 \text{ mg/m}^3$	NIOSH, 1994

Table 2. Physical and Chemical Properties of 1,1,2-TCA

Production and Uses

1,1,2-Trichloroethane is produced in the U.S., although not in California, by serial chlorination of ethylene. 1,1,2-TCA is a chemical intermediate in the synthesis of 1,1-dichloroethene, also known as vinylidene chloride (ATSDR, 1989). 1,1-Dichloroethene is used in the production of polyvinylidene chloride copolymers. These copolymers are used extensively in consumer products including flexible food film wraps, and carpet backing. U.S. EPA's Toxic Release Inventory showed no reported releases of 1,1,2-TCA in California in recent years (U.S. EPA, 2005; data extracted October 10, 2005).

ENVIRONMENTAL OCCURRENCE AND HUMAN EXPOSURE

Air

Because 1,1,2-TCA is listed as a federal hazardous air pollutant, the California Air Resources Board (ARB) identified it as a Toxic Air Contaminant in 1993 under AB2728 (ARB, 1996). 1,1,2-TCA is not one of the 75 Toxic Air Contaminants for which ARB reports monitoring data (ARB, 1998). The principal mechanism for removal of airborne 1,1,2-TCA is via reaction with the hydroxyl radical. In the atmosphere, 1,1,2-TCA is relatively reactive compared to other chlorinated ethanes. The most abundant products are hydrochloric acid, formyl chloride, phosgene, and chloroacetyl chloride (Spence and Hanst, 1978). The half-life of 1,1,2-TCA ranges from 24-50 days in unpolluted atmospheres to only a few days in polluted air (HSDB, 2005).

Soil

Published information on soil levels of 1,1,2-TCA in California is not available. 1,1,2-Trichloroethane was detected at 25 of 418 hazardous wastes sites on U.S. EPA's nationwide National Priority List (ATSDR, 1989). With soil sorption coefficient value estimates ranging from 83 to 209, 1,1,2-TCA is moderately to highly mobile in soils. When released to soil, 1,1,2-TCA is partially volatilized into air and partially leached into groundwater. Biodegradation proceeds very slowly, if at all (U.S. EPA, 2001b).

Water

Only 13 of 13,328 groundwater sources of public drinking water tested in California had measurable levels of 1,1,2-TCA. The highest of the 13 concentrations was 40 ppb and the lowest was 0.6 ppb. No 1,1,2-TCA was detected in any of the 755 surface water samples (DHS, 1999). Of 1,069 sites sampled in New Jersey, 8.7 percent had detectable 1,1,2-TCA (ATSDR, 1989). 1,1,2-TCA has been detected in surface water up to 0.6 ppb, tap water at 0.1 to 31 ppb, and in subsurface water up to 350,000 ppb (ATSDR, 1989). When released into surface water, 1,1,2-TCA primarily evaporates. Chemical losses by adsorption to sediment, aquatic hydrolysis, or biodegradation are not likely to be significant (U.S. EPA, 2001b).

METABOLISM AND PHARMACOKINETICS

Absorption

Due to its lipophilic, or fat-soluble, nature, 1,1,2-TCA is readily absorbed via inhalation and dermal routes (CPHF, 1987). There is evidence that 1,1,2-TCA is rapidly absorbed following inhalation by humans. Rats and mice were found to absorb more than half the respective oral doses of 70 mg/kg and 300 mg/kg (Mitoma *et al.*, 1985). Human retention of inhaled 1,1,2-TCA vapors would also be expected to be about 50 percent, based on these findings in rats and mice and on the inhalation uptake of similar solvents, including trichloroethylene, in beagle dogs and humans (Raabe, 1986,1988). 1,1,2-TCA is also absorbed through the skin of guinea pigs and mice (Jacobson *et al.*, 1977). Tsurata *et al.* (1975) measured the dermal absorption of 1,1,2-TCA after a 15-minute application of 0.5 mL to male ICR mice, producing an estimated absorption rate of about 130 nmole/min-cm² of skin. Based on the results, these authors calculated that humans could retain 13.9 mg/minute of 1,1,2-TCA in contact with both hands (Tsurata *et al.*, 1975).

Distribution

Following inhalation by mice of 1,000 parts per million (ppm) 1,1,2-TCA for one hour, the highest levels of the compound were detected in the fat. 1,1,2-TCA was also found in kidney, liver, blood, brain and heart in decreasing order of concentration. A single oral administration to rats and mice of 70 and 300 mg 1,1,2-TCA/kg, respectively, resulted in detectable levels in the livers of both species (Mitoma *et al.*, 1985). Based on the lipophilic nature of the chemical, the lack of an ionizable functional group, and the results of the dermal absorption studies, it can be concluded that 1,1,2-TCA will readily diffuse into the bloodstream and subsequently distribute to tissues of organs with high fat content (CPHF, 1987).

Metabolism

1,1,2-Trichloroethane is metabolized chiefly in the liver by the cytochrome P-450 system in the endoplasmic reticulum (CPHF, 1987). Mitoma *et al.* (1985) investigated the metabolic disposition of 1,1,2-TCA administered orally to rats and mice at 30 and 70 mg/kg-day, respectively, 5 days/week for 4 weeks, followed by a single dose of ¹⁴Clabeled 1,1,2-TCA. The authors determined that 81 percent of the total radiolabeled dose was metabolized in each species. They observed that 1,1,2-TCA undergoes glutathionedependent metabolism yielding S-carboxy-methylcysteine, thiodiacetic acid and chloroacetic acid (Mitoma *et al.*, 1985). Mazzullo *et al.* (1986) suggested that 1,1,2-TCA undergoes a cytochrome P-450-mediated oxidation process yielding a highly reactive free radical which could react directly with DNA or be oxidized to an electrophile (Mazzullo *et al.*, 1986; CPHF, 1987).

Excretion

Morgan *et al.* (1970) noted that 2.9 percent of the single-breath inhalation dose of 1,1,2-TCA was recovered via the exhaled air of a human volunteer during the first hour following exposure. Compared with 12 other common C_1 and C_2 chlorinated hydrocarbons, 1,1,2-TCA had the lowest excretion rate via exhalation.

Mitoma *et al.* (1985) collected urine, feces and the chamber air of mice and rats for 48 hours after a single oral dose of ¹⁴C-1,1,2-TCA. Radioactivity was measured in the urine and feces combined, identified as excreta. CO_2 and volatile metabolites in the chamber air were trapped separately and quantified. The animals were killed at 48 hours after dosing. The livers and kidneys were removed for analysis of ¹⁴C-protein binding and the radioactivity remaining in the carcass was measured. The authors observed that rats and mice excreted respectively 72 and 76 percent of an oral dose of 1,1,2-TCA via the urine. Table 2 shows the dose and percent of administered dose recovered in air, excreta and carcass as well as the total recovery.

Species	Dose (mg/kg)	Recovery	Expired Air	CO ₂ (a)	Excreta (b)	Carcass (c)	Metabolized (a)+(b)+(c)
	(ing/kg)		Percent of administered dose				
Rat	70	90.5	9.5	5.1	72	3.9	81
Mouse	300	88.1	6.8	3.1	76	2.3	81

Table 2. Excretion of 1,1,2-TCA (Mitoma et al., 1985)

TOXICOLOGY

Toxicological Effects in Animals

Acute Toxicity

White *et al.* (1985) administered 1,1,2-TCA via gavage to male and female CD-1 mice to evaluate acute lethal effects of the chemical. The authors administered one of seven doses ranging from 200 to 600 mg/kg to groups of eight mice of each sex. All mice receiving doses equal to or greater than 450 mg/kg became sedated within an hour. Most deaths occurred within 24 hours, and no deaths occurred after 48 hours. Necropsies revealed a dose-dependant irritation of the upper gastrointestinal tract. Fifty to 75 percent of the animals in each group had pale livers and up to 25 percent demonstrated lung damage of reddened or hemorrhagic areas. The LD₅₀ values were 378 mg/kg and 491 mg/kg respectively in male and female mice (White *et al.*, 1985).

The median lethal inhalation concentrations (LC₅₀s) for 6-hour exposures were 1654 ppm in rats (Bonnet, 1980, as cited in ATSDR, 1996) and 416 ppm in mice (Gradiski *et al.*,

1978, as cited in ATSDR, 1989) (both studies in French). These concentrations correspond to approximate dose levels of 1137 mg/kg and 458 mg/kg, respectively. For the rat dose level, we assumed 0.25 m^3 /d breathing rate, 0.25 kg body weight, and 50 percent retention. For the mouse dose, we assumed 0.04 m^3 /d breathing rate, 0.025 kg body weight, and 50 percent retention. The rodent strains were not stated.

In a study examining the effect of phenobarbital or 3-methylcholanthrene pretreatment on hepatotoxicity of trichloroethanes, Carlson (1973) observed deaths in 3 of 5 rats exposed to 2080 ppm 1,1,2-TCA for 2 hours (approximately 379 mg/kg) and no deaths among 5 rats exposed to 890 ppm for 2 hours (approximately 204 mg/kg). For the rat dose level, we assumed 0.25 m³/d breathing rate, 0.25 kg body weight, and 50 percent retention. Interestingly, 3-methylcholanthrene potentiated liver toxicity, but only at the higher 1,1,2-TCA concentration. At this higher exposure level, the author observed greater relative liver weights, decreased glucose-6-phosphatase, and increased SGPT and SGOT levels. The group treated with 1,1,2-TCA-only and the 1,1,2-TCA group pretreated with phenobarbital had no hepatic changes at either 1,1,2-TCA dose level.

A lethal dermal dose (LD_{50}) determined in rabbits was 3.73 mL/kg (Smyth *et al.*, 1969). Irritation and skin damage were identified in rabbits and guinea pigs, respectively.

Subchronic Toxicity

In a study to determine the potential of selected aliphatic hydrocarbon chemicals to initiate or promote neoplastic progression, Story *et al.* (1986) observed that rats gavaged with 69 mg/kg-d of 1,1,2-TCA 5 days per week for 7 weeks increased body weight at a rate 60 percent less than that of control animals. Male Osborne-Mendel rats, at 10 rats per group, were given partial hepatectomies and examined for initiation or promotion propensities. 1,1,2-Trichloroethane significantly increased the putative neoplastic promotion effect as measured via γ -glutamyltranspeptidase activity (Story *et al.*, 1986).

Male and female mice were exposed to plain water or 20, 200, or 2000 ppm of 1,1,2-TCA in water for 90 days (White *et al.*, 1985). Control groups for both sexes each had 48 mice and there were 32 mice in each of the exposed groups. The authors calculated time-weighted doses delivered based on fluid consumption and body weights (Table 3). There was a 30 percent decrease in water consumption in male mice at the highest dose level, with no corresponding decrease in females at the high dose level (White *et al.*, 1985).

1,1,2-TCA Concentration (ppm)	Male Dose (mg/kg-d)	Female Dose (mg/kg-d)
20	4.4	3.9
200	46	44
2000	305	384

Table 3. Ninety-day Oral Doses of 1,1,2-TCA in Mice (White et al., 1986)

Over 40 endpoints were monitored in these mice: organ weights, serum chemistry, hematological, and liver microsome parameters including glutathione levels and enzyme activities. For males, both liver and kidney weights were significantly decreased at the mid and upper dosage levels. For females, liver, spleen, and kidney weights increased significantly at the highest dose while brain weights decreased at the highest level and thymus weights decreased at the middle level only. Serum alkaline phosphatase in male mice was significantly elevated only at the highest dose level. In addition, liver glutathione levels in males were significantly decreased at the middle and highest dose level, but not the lowest dose level. Finally, hepatic cytochrome P-450 and aniline hydroxylase activity levels were decreased at the middle and highest, but not the lowest dose in females. While these observations indicate 1,1,2-TCA affects the liver, none of the measured liver parameters represents a serious toxicological effect. In addition, frank liver toxicity was not observed at higher dose levels in either sex of the study. The following table summarizes the subchronic NOAEL and LOAEL for the selected effects.

Effect Observed (sex)	LOAEL (mg/kg-d)	NOAEL (mg/kg-d)
Decreased liver and kidney weights (male)	46	4.4
Liver glutathione decrease (male)	46	4.4
Cytochrome P-450 decrease (female)	44	3.9
Aniline hydroxylase decrease (female)	44	3.9

Table 4. Significant Effect Levels in 90-day Mouse Study (White et al., 1986)

Genetic Toxicity

The results from genetic toxicity testing are not conclusive. Table 5 summarizes the reports from the literature and includes *in vitro* exposure tests (bacteria, yeast, fungi and mammalian cells) and *in vivo* tests. It is important to note that conducting tests *in vitro* with volatile chemicals requires some care to ensure that tests do not yield false negative results. If the chemical volatilizes so that the media concentration is very low, a negative result may occur with a genotoxic chemical. Therefore, the second column of the table indicates if the exposures were conducted in a sealed container such as a desiccator. Many of the non-mammalian species used in these tests lack the enzymes to convert chemicals to potentially reactive intermediates capable of mutating DNA, and mammalian activating enzymes can be added to the media during the exposure, as indicated in the third and fourth columns.

Test Species, Strains	Exposure Conditions	Activ With	ation Without	Reference	
Bacteria Reverse Mutation (Ames assay)					
<i>S. typhimurium</i> TA 1535	Unknown	-	-	Rannug et al., 1978	
<i>S. typhimurium</i> TA 98, 100, 1535	Desiccator assay, vapor exposure	-	-	Barber and Donish, 1982	
<i>S. typhimurium</i> TA 98, 100, 1535, 1537, 1538	Desiccator assay, vapor exposure	-	No data	Simmon <i>et al.</i> , 1977	
<i>S. typhimurium</i> TA 97, 98, 100, 1535 Desiccator assay vapor exposure		-	No data	Zeiger et al., 1992	
	Yeast Mu	tation Ass	ay		
S. cerevisiae D3	Suspension assay	-	-	Simmon et al., 1977	
S. cerevisiae D7	Unknown, but other chemicals positive	+	No data	Bronzetti et al., 1987	
	Fungi Chron	nosomal E	ffects		
Aspergillus nidulans	Sealed capped glass tube	+	No data	Crebelli et al., 1995	
	Mamm	alian Cells			
BALB/C-3T3 – viral transformation	Sealed chamber, vapor exposure	No data	+/-	Tu et al., 1985	
Rat - DNA repair	None	No data	+	Williams, 1983	
Mouse - DNA repair None		No data	-	Williams, 1983	
In Vivo Rodent Exposures					
Balb/c (single strand breaks in DNA)	IP injection	-	NA	Taninger et al., 1991	

Table 5. Genotoxicity of 1,1,2-TCA

Four publications reported 1,1,2-TCA to be negative in *Salmonella typhimurium*, and one paper reported it to be positive in *Saccharomyces*. 1,1,2-Trichloroethane induced DNA repair in hepatocytes isolated from rats but not from mice. This is in apparent contrast with the observation of hepatic tumors in mice but not in rats (see Carcinogenicity). Additionally, IARC (1999) summarized genetic test results showing that the chemical binds to DNA, RNA and caused strong S-phase induction but not unscheduled DNA synthesis in rodents tested *in vivo*. IARC noted that *in vitro* testing showed induction of DNA damage and micronuclei in human lymphocytes (IARC, 1999). The overall evidence regarding the genotoxic potential of 1,1,2-TCA is mixed, with some positive evidence for cytogenetic damage in mammalian cells and fungus, and mixed evidence for mutagenic activity in bacteria and yeast.

Developmental and Reproductive Toxicity

In a large comparative study of developmental effects of 55 chemicals in ICR/SIM mice, Seidenberg and coworkers administered a maternal dose of 350 mg/kg-d 1,1,2-TCA by gavage in corn oil on gestation days 8 through 12. Dams were allowed to deliver normally, but those that had not delivered by GD 21 or 22 were killed and their uteri were examined. Several growth and viability parameters were measured in the offspring as part of an evaluation of the Chernoff/Kavlock developmental toxicity screen. No significant developmental toxic effects of 1,1,2-TCA were observed (Seidenberg *et al.*, 1986).

Immunotoxicity

Sanders and coworkers assessed the immunological effect of 1,1,2-TCA on CD-1 mice following oral exposure for 14 or 90 days. For the 14-day range-finding study, male rodents were exposed to 3.8 or 38 mg/kg. The authors observed no alterations in either humoral or cell-mediated immune status. For the 90-day drinking water exposure, males received 4.4, 46, or 305 mg/kg-d and females received 3.9, 44, and 384 mg/kg-d. Cell mediated immunity was unchanged for both sexes. Humoral immune status, determined by hemagglutination titers, was significantly depressed at 46 mg/kg-d, but not at 4.4 mg/kg-d. Therefore, these values were selected as the LOAEL and NOAEL for this study, respectively (Sanders *et al.*, 1985).

Neurotoxicity

Kallman and associates (1983) evaluated five halogenated hydrocarbons including 1,1,2-TCA to determine their thresholds for induction of conditioned taste aversion in male CD-1 mice (a sensitive test for adverse neurological effects). The authors administered 3, 10, 30, 100, or 300 mg/kg 1,1,2-TCA to mice once via gavage in acute trials, and once daily for seven consecutive days, with seven mice per dose. When paired with saccharin ingestion, a dose-related aversion to saccharin in drinking water was observed at the two highest levels. Accordingly, the LOAEL for 1,1,2-TCA in this study is 100 mg/kg-d and the NOAEL is 30 mg/kg-d (Kallman *et al.*, 1983). Note that although this is a behavioral test that can be sensitive to neurological (aversive) effects, any adverse effect including gastrointestinal discomfort is capable of inducing conditioned aversion. Therefore this test response is more generally classified as acute toxicity rather than neurotoxicity.

Chronic Toxicity

NCI (1978) studied the effects of technical grade 1,1,2-TCA administered orally in corn oil on rats and mice. Groups of 50 male and female Osborne-Mendel rats were treated via gavage five days per week for 78 weeks. Rats of both sexes received 35 or 50 mg/kg-d of 1,1,2-TCA for 20 weeks. The respective doses were then increased to 50 or 100 mg/kg-d for the remaining 58 weeks. The time-weighted average doses were 46 and 92 mg/kg-d, respectively, for the low and high dose groups for both sexes. The rats were observed for 35 weeks post-treatment.

In the same study, groups of 50 male and female $B6C3F_1$ mice were treated via gavage five days per week for 78 weeks. Mice of both sexes received 150 or 300 mg/kg-d of 1,1,2-TCA for eight weeks. The respective doses were then increased to 200 or 400 mg/kg-d for the remaining 58 weeks. The time-weighted average doses were 195 and 390 mg/kg-d, respectively, for the low and high dose groups for both sexes. The mice were observed for 13 weeks post-treatment.

For both rats and mice, two control groups of 20 animals of each sex received either corn oil alone (vehicle control) or remained untreated. The NCI protocol was designed primarily to test the carcinogenic potential of a chemical, but non-neoplastic lesions were evaluated by a pathologist examining histological sections of the organs of animals by light microscopy. No significant increases in non-neoplastic lesions were reported in the organ systems examined, including liver, in either rats or mice. Therefore the highest doses, 92 mg/kg-d for rats and 390 mg/kg-day for mice, are determined to be chronic NOAELs for noncancer effects in these NCI studies.

Carcinogenicity

Oral Exposure

Only one report examining the carcinogenicity of 1,1,2-TCA in animals from long-term oral exposure was found (NCI, 1978). The details of dosing of rats and mice are described in the *Chronic Toxicity* subsection. Neither sex of Osborne-Mendel rats showed an increased incidence of any type of neoplasm at either of the two doses. In both sexes of $B6C3F_1$ mice in both treatment groups, the incidence of hepatocellular carcinomas was significantly increased above the incidence in control animals. The increases in liver tumor incidence were also dose-dependent, showing a monotonic increase in both sexes. Positive trends with increasing dose were observed for adrenal pheochromocytomas in both male and female mice, although a statistically significant increase over the incidence in control animals was only observed among female mice in the high dose group.

Hepatocellular carcinoma is a neoplasm arising from the most abundant cell in the liver, the hepatocyte. Some strains of mice such as the C57Bl/6 have very high spontaneous incidence rates of this particular tumor. In contrast, C3H mice have a much lower spontaneous rate of hepatocellular carcinoma. The B6C3H is the first generation (F_1) of a cross of the C57Bl/6 and C3H inbred mouse strains. B6C3H mice were selected for conducting carcinogenicity bioassays like that for 1,1,2-TCA because the use of an F_1 hybrid was thought to balance the sensitivity to carcinogens with a moderate background level of hepatocellular carcinoma.

Pheochromocytoma is a neoplasm stemming from a chromaffin cell in the medulla of the adrenal gland. These cells have vesicles that contain the catecholamines epinephrine and norepinephrine. The catecholamines can be oxidized and polymerized after reacting with potassium bichromate (chromaffin reaction). Chromaffin cells appear to be filled with fine brown granules after adrenal tissue sections are stained with potassium bichromate.

In the NCI mouse studies, the authors observed that 1,1,2-TCA produced significant increases in liver tumors, to a relatively high incidence, in a dose-dependent manner in chronic exposure to both male and female mice. Please see Table 6.

	Control (Untreated)	Control (Vehicle)	Low Dose	High Dose
Male	2/17 (12%)	2/20 (10%)	18/49 (37%)	37/49 (76%)
Female	2/20 (10%)	-	16/48 (33%)	40/45 (89%)

Table 6. Hepatocellular Carcinoma Incidence in Mice (NCI, 1978)

The interpretation of the NCI (1978) rat bioassays is limited by the relatively short treatment duration of 78 weeks. Short treatment duration is generally a limitation of negative studies. When less-than-lifetime studies show positive carcinogenicity findings – as this one does in mice – concern is enhanced.

An uncertainty in the NCI studies is the potential presence of toxic impurities in the technical grade 1,1,2-TCA used as the test substance. Three purity tests were performed by gas-liquid chromatography and infrared spectroscopy on the batch of chemical used in these studies, with a determination of 99.2 percent purity at the beginning of the study, and two further determinations of 90.8 percent and 95.4 percent, seven and 13 months into the studies, respectively. The major impurities were not identified in the report (NCI, 1978). The NCI studies were conducted before the implementation of current Good Laboratory Practices (GLP), although no specific evidence casts doubt on the findings (ATSDR, 1989).

Subcutaneous Exposure

Norpoth *et al.* (1988) investigated carcinogenicity in male and female Sprague-Dawley rats subcutaneously injected with 15.4 or 46.8 μ mole of 1,1,2-TCA in DMSO once a week for two years. The authors observed an increased incidence of sarcomas, mostly localized on the extremities. The incidence was not significant when compared with vehicle controls, but was significant when compared with untreated controls. No sarcomas were reported in the untreated controls.

Transgenic Mouse Studies

Lifetime rodent bioassays are expensive and time-consuming. There have been a number of efforts to find alternatives that could be used to identify carcinogens more rapidly. Yamamoto *et al.* (1998) developed a transgenic strain of C57BL/6J mice having five or six copies of the human c-Ha-*ras* gene integrated into its genome. The F_1 generation from a cross of males of this transgenic genotype with normal female BALB/cByJ were exposed to a variety of chemicals of known carcinogenic potential for 26 weeks or less and examined for tumors. Validation studies have been conducted to determine if these

mice will predict the carcinogenic human potential of a chemical such as animal bioassays using $B6C3F_1$ mice and Fischer rats. 1,1,2-TCA was selected as a chemical that, according to the authors, was carcinogenic but not mutagenic. In this study, 1,1,2-TCA induced no tumors, despite doses of chemical that were high enough to induce evidence of liver toxicity (Yamamoto *et al.*, 1998).

Toxicological Effects in Humans

Acute Toxicity

The human data available to evaluate the toxicity of 1,1,2-TCA are "very limited" (RAIS, 2002). The chemical is rapidly absorbed by the body and has a narcotic effect at "low" concentrations. Exposure to 1,1,2-TCA is irritating to eyes and the mucous membranes of the respiratory tract (RAIS, 2002).

Wahlberg *et al.* (1984) observed that a five-minute exposure of 698 mg 1,1,2-TCA/cm² to the forearm skin of a healthy male subject resulted in erythema as assessed by laser Doppler flowmetry. The subject reported a stinging or burning sensation (Wahlberg, 1984).

Subchronic Toxicity

No human studies were found.

Genetic Toxicity

IARC (1999) reports that 1,1,2-TCA induced DNA damage and micronuclei in *in vitro* testing of human lymphocytes.

Developmental and Reproductive Toxicity

No human studies were found.

Immunotoxicity

No human studies were found.

Neurotoxicity

No human studies were found.

Chronic Toxicity

No human studies were found.

Carcinogenicity

Dosemeci *et al.* (1999) examined potential risk of renal cell carcinoma from occupational exposures to chlorinated aliphatic hydrocarbons, with an emphasis on gender differences. Among women, significantly elevated relative risks of renal cell carcinoma were associated with exposure to "all organic solvents" (odds ratio [OR] = 2.3, confidence interval [CI] = 1.3-4.2), "all chlorinated aliphatic hydrocarbons combined" (OR = 2.1, CI = 1.1-3.9), and to trichloroethylene alone (OR = 2.0, CI = 1.0-4.0), but not to 1,1,2-TCA alone. None of the comparisons were significant for men. Exposure levels were low for individual solvents, so statistical power was limited

DOSE-RESPONSE ASSESSMENT

Noncarcinogenic Effects

In humans, exposure to 1,1,2-TCA can cause irritation to skin and mucous membranes and has a narcotic effect. In experimental animals, there were no non-neoplastic adverse effects reported in the only chronic oral exposure study identified (NCI, 1978). The 90day drinking water study in mice (White *et al.*, 1985) described in the Subchronic Toxicity subsection provides the best study on which to estimate a health-protective concentration for drinking water for non-cancer effects because the nature of the exposure is the closest to the human drinking water exposure conditions. In addition, the effect level identified for this study is the lowest of the available studies. The liver was clearly affected by 1,1,2-TCA in both male and female mice at the middle and highest but not the lowest dose. A LOAEL of 44 mg/kg-d and NOAEL of 3.9 mg/kg-d were selected based on liver effects in female mice. For comparison, the chronic time-weighted average doses in the NCI (1978) mouse study were 195 and 390 mg/kg-d, respectively, for the low and high dose groups, for both sexes.

Carcinogenic Effects

Based upon the increased incidence of liver tumors observed in both male and female mice treated with two doses of 1,1,2-TCA, and on the development of pheochromocytomas in both sexes of mice, this compound should be treated as a potential human carcinogen. The mechanism by which these tumors are induced in mice is not presently understood. Although not all tests for genotoxicity were positive, some evidence for genetic damage (*e.g.*, cytogenetic damage) may point to the involvement of this mechanism in the tumors induced by 1,1,2-TCA.

1,1,2-Trichloroethane appears on the Proposition 65 list of chemicals known to cause cancer (as vinyl trichloride). A Proposition 65 No Significant Risk Level of 10 micrograms/day has been promulgated based upon the liver tumor findings in female mice, which appear to be slightly more sensitive than male mice to the carcinogenic effects of 1,1,2-TCA, resulting in a calculated human oral cancer potency estimate of 0.072 (mg/kg-d)⁻¹ (OEHHA, 1992, 1999). U.S. EPA (2004) has estimated a carcinogenic

potency of 0.057 (mg/kg-d)⁻¹ for 1,1,2-TCA based on the male mouse hepatocellular carcinoma data from NCI (1978). Recently, OEHHA re-evaluated the cancer potency factor for 1,1,2-trichloroethane and concurred with the U.S. EPA carcinogenic potency of 0.057 (mg/kg-d)⁻¹ (OEHHA, 2005).

CALCULATION OF PHG

Calculations of concentrations of chemical contaminants in drinking water associated with negligible risks for carcinogens or noncarcinogens must take into account the toxicity of the chemical and the potential exposure of individuals using the water.

Exposure Considerations

The value of 2 L/day for water consumption is a traditional default value for an adult, nominally representing all uses of water, including that added to food and used for coffee or tea. Inhalation of volatile organic chemicals released from water used in the household may result in additional human exposure. Calculations of equivalent exposures from other household uses of drinking water with the CalTOX multimedia exposure program yield an amount roughly equal to the drinking water contribution for small, volatile halogenated hydrocarbons such as 1,1,2-TCA. Thus, we have added an exposure equivalent to drinking two L/day to account for exposures from bathing or showering. This total exposure of four Leq/day accounts for exposures to a toxicant from all household uses of drinking water.

Noncarcinogenic Effects

Calculation of a public health-protective concentration (C, in mg/L) for a chemical in drinking water for noncarcinogenic endpoints follows the general equation:

$$C = \frac{NOAEL \times BW \times RSC}{UF \times WC} = mg/L$$

where,

NOAEL	=	no-observed-adverse-effect-level (3.9 mg/kg-d);
BW	=	adult body weight (70 kg);
RSC	=	relative source contribution (80 percent);
UF	=	uncertainty factor of 10,000, which includes factors of 10 for inter- species extrapolation, 10 for subchronic to chronic extrapolation, 10 for potentially sensitive human subpopulations, and 10 for potential carcinogenicity;
WC	=	equivalent daily water consumption/contact rate (4 L_{eq}/d).

The 90-day drinking water study in mice (White *et al.*, 1985) was used to determine a LOAEL of 44 mg/kg-d and a NOAEL of 3.9 mg/kg-d based on liver effects in female mice. Using these parameter values, a health-protective value (C) is calculated as follows:

C =
$$3.9 \text{ mg/kg-d x 70 kg x 0.8} = 0.005 \text{ mg/L} = 5 \text{ ppb}$$

10,000 x 4 L_{eq}/d

Based on this calculation, the noncancer health-protective value for 1,1,2-TCA in drinking water is 5 ppb.

Carcinogenic Effects

Calculation of carcinogenic potency utilizes the following equation:

$$C = \frac{R \times BW}{CPF \times L/day} = mg/L$$

where,

R	=	<i>de minimis</i> level for lifetime excess individual cancer risk (a default of 10^{-6});
BW	=	body weight for an adult male (70 kg);
CPF	=	the cancer potency factor or the oral potency slope; the CPF for $1,1,2$ -TCA is 0.057 (mg/kg-day) ⁻¹ ;
L/day	=	volume of drinking water consumed by an adult; the default is 2 L/d, but multiroute exposures should be considered as discussed above.

Therefore,

C =
$$\frac{(1 \times 10^{-6}) \times 70 \text{ kg}}{(0.057) \times 4 \text{ L}_{eq}/\text{day}}$$
 = 0.000307 mg/L or 0.3 ppb (rounded)

Based on the above considerations, the estimated health-protective concentration for carcinogenicity is much lower than for non-cancer effects. We calculate a PHG value of 0.3 ppb for 1,1,2-TCA based on a cancer risk level of one cancer in a million people exposed for a lifetime to the chemical in their drinking water. The equivalent concentration in drinking water at a risk level of one in 10,000 would be 30 ppb, and at 1 in 100,000 would be 3 ppb.

RISK CHARACTERIZATION

The PHG value was based on carcinogenic, rather than non-cancer effects, for two reasons. First, the cancer-based PHG is lower than the non-cancer by a factor of over 10, and thus is more health protective. Second, the non-cancer PHG calculation had a greater than typical amount of uncertainty associated with its derivation, to the point of an uncertainty divisor of 10,000 when a factor for uncertainty about carcinogenicity is added. Other sources of uncertainty in the development of the health-protective value for non-cancer endpoints for 1,1,2-TCA in drinking water are also the general issues of uncertainty in many risk assessments, particularly mode of action and inter- and intraspecies extrapolations. The subchronic duration of the critical noncancer study provides additional uncertainty in this case. Uncertainty in the relative source contribution (RSC) is relatively minimal for this solvent, because there are few if any other common sources of 1,1,2-TCA.

The carcinogenic effect-based PHG also includes uncertainty, however. The frank carcinogenicity was only found in one animal species, albeit both sexes and two tissues of origin. 1,1,2-Trichloroethane was shown to result in statistically significant increases in hepatocellular carcinoma at two doses and pheochromocytoma at the high dose in $B6C3F_1$ mice of both sexes. Concern for a carcinogenic endpoint is generally enhanced by a positive cancer data in more than one species, or from replicate positive studies in the same species. However, in the case of 1,1,2-TCA, concern is also increased by evidence of a dose-related increase in the carcinogenic response and by the magnitude of the response. The fact that 1,1,2-TCA was positive in only one species resulted in U.S. EPA classifying it as a "possible" (group C), rather than "probable," human carcinogen. The International Agency for Research on Cancer has classified 1,1,2-TCA as "not classifiable as to its carcinogenicity to humans" (Group 3) based upon limited evidence in experimental animals and the absence of epidemiological evidence (IARC, 1999). However, IARC notes the ability of 1,1,2-TCA to bind to DNA, RNA and protein, cause strong S-phase induction in rodents in vivo, and induce DNA damage and micronuclei in human lymphocytes and cell transformation in BALB/c-3T3 cells in vitro (IARC, 1999). Such effects can indicate potential carcinogenicity.

1,1,2-TCA did not produce a tumor response following relatively short-term exposures when tested in a transgenic strain of C57BL/6J mice with five or six integrated copies of the human c-Ha-*ras* gene. Other hepatocarcinogens have caused liver tumors in these transgenic mice. Further complicating interpretation of the available data, 1,1,2-TCA increased putative preneoplastic lesions in rats in a study examining tumor promotion properties (Story *et al.*, 1986; see Subchronic Toxicity section).

Confirmatory studies, such as human epidemiological corroboration and *in vitro* genetic toxicity testing, were generally absent, negative or inconclusive, although the number of tests was relatively small. OEHHA considers it prudent to assume carcinogenicity of 1,1,2-TCA for this risk assessment purpose, which is to estimate a concentration of the solvent in drinking water that is unlikely to provide any toxic hazard after a lifetime of exposure. OEHHA considers that the PHG value of 0.3 ppb is adequately health protective for potential sensitive subpopulations, including infants, children, and the elderly.

OTHER REGULATORY STANDARDS

U.S. EPA proposed a federal Maximum Contaminant Level Goal (MCLG) of 0.003 mg/L and an MCL of 0.005 mg/L for 1,1,2-TCA in 1990 (U.S. EPA, 1990) and finalized these two criteria in 1992 (U.S. EPA, 1992). The federal MCLG is computed using a NOAEL from a 90-day drinking water exposure study in mice (Sanders, 1985; White, 1985) and a safety factor of 10,000, whereas the MCL is set at the practical quantitation limit (U.S. EPA, 2002). California subsequently promulgated an MCL for 1,1,2-TCA of 0.005 mg/L (DHS, 2002), consistent with the U.S. EPA MCL. U.S. EPA recently reviewed their primary drinking water regulations and concluded that it might be possible to decrease the MCL from the present value of 0.005 mg/L based on methodological improvements since the original regulation was promulgated (U.S. EPA, 2002). However, the U.S. EPA concluded that a revision to the MCL is not warranted at this time, because 1,1,2-TCA is very rarely detected in drinking water at concentrations exceeding the MCLG.

The available regulatory standards and guidelines are summarized in Table 7, below.

Agency	Standard or Criterion	Level	Comment
ACGIH	TLV-TWA	10 ppm	8-hr work day
OSHA	PEL (permissible exposure limit)	10 ppm	8-hr work day
U.S. EPA	MCL (maximum contaminant level)	0.005 mg/L	Based on historical value for practical quantitation limit
U.S. EPA	MCLG (maximum contaminant level goal)	0.003 mg/L	Based on non-cancer endpoint
U.S. EPA	Ten-day Health Advisory	0.40 mg/L	
CDHS	MCL (maximum contaminant level)	0.005 mg/L	Based on U.S. EPA MCL

Table 7. Selected Guidelines and Regulations for 1,1,2-Trichloroethane
--

Table adapted from ATSDR (1989), NIOSH (1994), U.S. EPA (2001), DHS (2003).

REFERENCES

ARB (1996). Toxic Air Contaminant Identification List - June 1996. California Air Resources Board Website. Accessed at: http://www.arb.ca.gov/toxics/tac/taclist.htm.

ARB (1998). California Ambient Air Quality Data 1980-1997. California Air Resources Board, Sacramento, California.

ATSDR (1989). Toxicological Profile for 1,1,2-Trichloroethane. December, ATSDR/TP-89-12. Prepared by Clement Associates under Contract No. 205-88-0608, Agency for Toxic Substances and Disease Registry, U.S. Department of Health and Human Services (USDHHS), Centers for Disease Control (CDC), in collaboration with the U.S. EPA, with technical editing and document preparation by Oak Ridge National Laboratory. Atlanta, Georgia.

Barber ED, Donish WH (1982). An exposure system for quantitative measurements of the microbial mutagenicity of volatile liquids. In: Genotoxic Effects of Airborne Agents. Tice RR, Costa DL, Schaich KM, eds. Plenum Press, New York.

Bonnet P, Francin JM, Gradiski D (1980). Determination of the median lethal concentration of the principal chlorinated aliphatic solvents in rats. Arch Mal Prof Med Trav Secur Soc 41:317-21.

Bronzetti G, Galli A, Vellosi R, Rossi F, Morichetti E, Del Carratore R (1987). Genetic activity of chlorinated ethanes. IXth meeting of the European Association for Cancer Research, Helsinki, Finland. Cancer Clin Oncol 23:1737-8.

Carlson GP (1973). Effect of phenobarbital and 3-methylcholanthrene pretreatment on the hepatotoxicity of 1,1,1-trichloroethane and 1,1,2-trichloroethane. Life Sci 13:67-73.

Cleseri LS, Greenberg AE, Trussel AR, eds. (1989). Volatile Organics. In: Standard Methods for the Examination of Water and Wastewater. American Public Health Association, Washington, DC.

CPHF (1987). Health risk assessment of 1,1,2-trichloroethane (1,1,2-TCA) in California drinking water, Draft 1. Risk Assessment Group, Dept. Environmental Toxicology, University of California at Davis, for California Public Health Foundation, State of California contract number 84-84571.

Crebelli R, Andreoli C, Carere A, Conti L, Crochi B, Cotta-Ramusino M, Benigni R (1995). Toxicology of halogenated aliphatic hydrocarbons: Structural and molecular determinants for the disturbance of chromosome segregation and the induction of lipid peroxidation. Chem-Biol Interact 98:113-29.

DHS (1988). Notice of Proposed Rulemaking. Maximum contaminant levels for 1,1,2trichloroethane, thiobencarb, molinate, bentazon, atrazine, and simazine in drinking water (R-39-88). California Department of Health Services (DHS), Health and Welfare Agency, Sacramento, California.

DHS (1999). Drinking Water Monitoring Data 1984-1998. Annual Status Report. California Department of Health Services, Sacramento, California.

DHS (2003). Extract from Title 22, California Code of Regulation, Div 4, Chap 15, Domestic Water Quality and Monitoring, Article 5.5, Primary Standards – Organic Chemicals, Section 64444, Maximum contaminant levels – organic chemicals. Accessed at: <u>http://www.dhs.ca.gov/ps/ddwem/chemicals/mcl/regextract.pdf</u>.

Dosemeci M, Cocco P, Chow W (1999). Gender differences in risk of renal cell carcinoma and occupational exposures to chlorinated aliphatic hydrocarbons. Am J Indust Med 36:54-9.

Gradiski D, Bonnet P, Raoult G (1978). Comparative acute inhalation toxicity of the principal chlorinated aliphatic solvents. Arch Mal Prof Med Trav Secur Soc 39:249-57.

HSDB (2005). 1,1,2-Trichloroethane (CASRN: 79-00-5). Hazardous Substances Data Bank, National Library of Medicine, Toxnet toxicology data network. Accessed at: http://toxnet.nlm.nih.gov/cgi-bin/sis/search.

IARC (1999). Summary for monograph on 1,1,2-trichloroethane. Accessed at: <u>http://www-cie.iarc.fr/htdocs/monographs/vol71/049-trichloret.html</u>.

Jakobson I, Holmberg B, Wahlberg JE (1977). Variations in the blood concentration of 1,1,2-trichloroethane by absorption and other routes of administration in the guinea pig. Acta Pharmacol Toxicol (Copenh) 41:497-506.

Kallman MJ, Lynch MR, Landauer MR (1983). Taste aversions to several halogenated hydrocarbons. Neurobehav Toxicol Teratol 5:23-7.

Mazzullo M, Colacci A, Grilli S, Prodi G, Arfellini G (1986). 1,1,2-Trichloroethane: evidence of genotoxicity from short-term tests. Jpn J Canc Res 77:532-9.

McKone TE (1987). Human exposure to volatile organic compounds in household tap water - the indoor inhalation pathway. Environ Sci Technol 21:1194-201.

Mitoma C, Tyson CA, Riccio ES (1985). Investigation of the species sensitivity and mechanism of carcinogenicity of halogenated hydrocarbons. EPA/OTS, Document #40-842-8424225.

Morgan A, Black A, Belcher DR (1970). The excretion in breath of some aliphatic halogenated hydrocarbons following administration by inhalation. Ann Occup Hyg 13:219-33.

NCI (1978). Bioassay of 1,1,2-trichloroethane for possible carcinogenicity. National Cancer Institute, Carcinogenesis Technical Report Series Number 74. NCI-CG-TR-74. PB-28337.

NIOSH (1994). Pocket Guide to Chemical Hazards. U.S. Department of Health and Human Services, pp. 314-5.

Norpoth K, Heger M, Muller G, Mohtashmipur E, Kemena A, Wittig C (1988). Investigations on metabolism and carcinogenicity of 1,1,2-trichloroethane. J Cancer Res Clin Oncol 114 (2):158-62.

OEHHA (1992). Expedited cancer potency values and proposed regulatory levels for certain Proposition 65 carcinogens. Reproductive and Cancer Hazard Assessment Section, Office of Environmental Health Hazard Assessment, April 1992.

OEHHA (1993). Review of U.S. EPA's final rule of phase V chemicals. Memorandum dated March 15, 1993, from J. Brown, Pesticide and Environmental Toxicology Unit, Office of Environmental Health Hazard Assessment, to Alexis Milea, Chemical Standards and Technology Unit, California Department of Health Services, Sacramento, California.

OEHHA (1994). California Cancer Potency Factors. Memorandum from David Siegel to the Standards and Criteria Workgroup. Office of Environmental Health Hazard Assessment, California Environmental Protection Agency, Sacramento, California.

OEHHA (2005). Air Toxics Hot Spots Program Risk Assessment Guidelines; Part II, Technical Support Document For Describing Available Cancer Potency Factors, May 2005. Office of Environmental Health Hazard Assessment, California Environmental Protection Agency, Sacramento, California. Accessed at: http://www.oehha.ca.gov/air/hot_spots/pdf/May2005Hotspots.pdf.

Raabe OG (1986). Inhalation uptake of selected chemical vapors at trace levels. Final Report to the California Air Resources Board, CARB contract No. A3-132-33. University of California, Davis.

Raabe OG (1988). Retention and metabolism of toxics; inhalation uptake of xenobiotic vapors by people. Final Report to the California Air Resources Board, CARB contract No. A5-155-33. University of California, Davis.

RAIS (2002). Toxicity Profile, RAGs, A Format for 1,1,2-Trichloroethane - CAS Number 79005. Risk Assessment Information System, Oak Ridge National Laboratory. Accessed at http://risk.lsd.ornl.gov/tox/profiles/1_1_2_trichloroethane_ragsa.shtml.

Rannug U, Sundvall A, Ramel C (1978). Mutagenic effect of 1,2-dichloroethane on *salmonella typhimurium*. 1. Activation through conjugation with glutathione *in vitro*. Chem Biol Interact 20:1-16.

Sanders VM, White KL, Jr, Shopp GM, Jr, Munson AE (1985). Humoral and cellmediated immune status of mice exposed to 1,1,2-trichloroethane. Drug Chem Toxicol 8:357-72.

Seidenberg JM, Anderson DG, Becker RA (1986). Validation of an *in vivo* developmental toxicity screen in the mouse. Teratog Carcinog Mutagen 6:361-74.

Simmon VF, Kauhanen K, Tardiff RG (1977). Mutagenic activity of chemicals identified in drinking water. Dev Toxicol Environ Sci 2:249-58.

Smyth HF, Jr, Carpenter CP, Weil CS, Pozzani UC, Striegel JA, Nycum JS (1969). Range-finding toxicity data: List VII. Am Ind Hyg Assoc J 30:470-6.

Spence J, Hanst P (1978). Oxidation of chlorinated ethanes. J Air Pollut Cont Assoc 28:250-3.

Story DL, Meierhenry EF, Tyson CA, Milman HA (1986). Differences in rat liver enzyme-altered foci produced by chlorinated aliphatics and phenobarbital. Toxicol Ind Health 2:351-62.

Taningher M, Parodi S, Grilli S, Colacci A, Mazzullo M, Bordone R, Santi L (1991). Lack of correlation between alkaline DNA fragmentation and DNA covalent binding induced by polychloroethanes after in vivo administration: Problems related to the assessment of a carcinogenic hazard. Canc Detect Prev 15:35-40.

Tsurata H (1975). Percutaneous absorption of organic solvents. Ind Health 13:227-236.

Tu AS, Murray TA, Hatch KM, Sivak A, Milman HA (1985). In vitro transformation of BALB/c-3T3 cells by chlorinated ethanes and ethylenes. Canc Lett 28:85-92.

U.S. EPA (1990). National Primary Drinking Water Regulations, Synthetic Organic Chemicals and Inorganic Chemical, Proposed Rule. Fed Reg July 25, 1990, p. 30406.

U.S. EPA (1992). National Primary Drinking Water Regulations, Synthetic Organic Chemicals and Inorganic Chemical, Final Rule. Fed Reg July 17, 1992, p. 31795.

U.S. EPA (2001a). Current drinking water standards, National Primary Drinking Water Regulations. Office of Water, U.S. Environmental Protection Agency. Accessed at: www.epa.gov/safewater/mcl.html.

U.S. EPA (2001b). Technical fact sheet on 1,1,2-trichloroethane. National Primary Drinking Water Regulations. Office of Water, U.S. Environmental Protection Agency. Accessed at: www.epa.gov/safewater/dwh/t-voc/1,1,2-trichloroethane-tric.html.

U.S. EPA (2002). National Primary Drinking Water Regulations, Announcement of the results of EPA's review of existing drinking water standards and request for public comment. Fed Reg 67(74):19029-19090 (April 17, 2002).

U.S. EPA (2004). 1,1,2-Trichloroethane (carcinogenicity assessment posted 02/01/1994). Integrated Risk Information System, U.S. Environmental Protection Agency, Washington, DC. Accessed at: http://www.epa.gov/iris/subst/index.html.

U.S. EPA (2005). Toxic Release Inventory. U.S. Environmental Protection Agency, Washington, DC. Accessed at: http://www.epa.gov/enviro/html/tris/adhoc.html.

Wahlberg JE (1984). Erythema-inducing effects of solvents following epicutaneous administration to man - studied by laser Doppler flowmetry. Scand J Work Environ Health 10:159-62.

White KL, Jr, Sanders VM, Barnes DW, Shopp GM, Jr, Munson AE (1985). Toxicology of 1,1,2-trichloroethane in the mouse. Drug Chem Toxicol 8:333-55.

Williams GM (1983). DNA repair tests of 11 chlorinated hydrocarbon analogs. Final Report, EPA contract. NTIS/OTS 0509403.

Yamamoto S, Urano K, Nomura T (1998). Validation of transgenic mice harboring the human prototype c-Ha-ras gene as a bioassay model for rapid carcinogenicity testing. Toxicol Lett 102-103:473-8.

Zeiger E (1987). Carcinogenicity of mutagens: Predictive capability of the *Salmonella* mutagenesis assay for rodent carcinogenicity. Canc Res 47:1287-96.